

## Medicinal plants with anti-SARS-CoV activity repurposing for treatment of COVID-19 infection: A systematic review and meta-analysis

OLUWAKEMI EBENEZER<sup>1\*</sup>  
OLUSOLA BODEDE<sup>2</sup>  
PAUL AWOLADE<sup>3</sup>  
MARYAM A. JORDAAN<sup>1</sup>  
ROPO E. OGUNSAKIN<sup>4</sup>  
MICHAEL SHAPI<sup>1</sup>

<sup>1</sup> Faculty of Natural Science, Department of Chemistry, Mangosuthu University of Technology, 511 Mangosuthu Highway Durban, 4000, South Africa

<sup>2</sup> Department of Agriculture and Animal Health, University of South Africa Florida Campus, Florida, 1710 South Africa

<sup>3</sup> School of Chemistry and Physics University of KwaZulu-Natal P/Bag X54001, Westville, Durban South Africa

<sup>4</sup> Discipline of Public Health Medicine College of Health Sciences University of KwaZulu Natal, Durban 4001, South Africa

The novel SARS-CoV-2 (severe acute respiratory syndrome coronavirus) has emerged as a significant threat to public health with startling drawbacks in all sectors globally. This study investigates the practicality of some medicinal plants for SARS-CoV-2 therapy using a systematic review and meta-analysis of their reported SARS-CoV-1 inhibitory potencies. Relevant data were systematically gathered from three databases, *viz.*, Web of Science, PubMed and Scopus. The information obtained included botanical information, extraction method and extracts concentrations, as well as the proposed mechanisms. Fourteen articles describing 30 different plants met our eligibility criteria. Random effects model and subgroup analysis were applied to investigate heterogeneity. According to subgroup analysis, the substantial heterogeneity of the estimated mean based on the  $IC_{50}$  values reporting the most potent anti-SARS-CoV 3C-like protease (3CLpro) inhibitors (10.07 %,  $p < 0.0001$ ), was significantly higher compared to the most active anti-SARS-CoV papain-like protease (PLpro) inhibitors (6.12 %,  $p < 0.0001$ ). More importantly, the literature analysis revealed that fruit extracts of *Rheum palmatum* L. and the compound cryptotanshinone isolated from the root of *Salvia miltiorrhiza* ( $IC_{50} = 0.8 \pm 0.2 \mu\text{mol L}^{-1}$ ) were excellent candidates for anti-SARS-CoV targeting PLpro. Meanwhile, iguesterin ( $IC_{50} = 2.6 \pm 0.6 \mu\text{mol L}^{-1}$ ) isolated from the bark of *Tripterygium regelii* emerged as the most excellent candidate for anti-SARS-CoV targeting 3CLpro. The present systematic review and meta-analysis provide valuable and comprehensive information about potential medicinal plants for SARS-CoV-2 inhibition. The chemotypes identified herein can be adopted as a starting point for developing new drugs to contain the novel virus.

**Keywords:** COVID-19 main protease, inhibition, medicinal plants, systematic review, meta-analysis

Accepted September 14, 2021  
Published online September 15, 2021

\* Correspondence; e-mail: ebenezer.oluwakemi@mut.ac.za

## INTRODUCTION

The severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) has been identified as the causative agent for the novel pneumonia-type coronavirus disease 2019 (COVID-19) (1, 2). The outbreak of this disease remains among public health emergencies of international concern as earlier declared by the World Health Organisation (3). As of August 16<sup>th</sup> 2021, about 222 countries have been affected by this coronavirus, and there were ~209 million confirmed cases and ~ 4.4 million deaths have been recorded globally. The symptoms of COVID-19 include cough, fever, fatigue, myalgia and dyspnoea, with the less common manifestation of runny nose, headache, nasal congestion, sore throat, and diarrhoea (1, 4). Severe conditions such as pneumonitis, acute respiratory distress syndrome (ARDS), respiratory arrest, inflammatory-induced lung injury, sepsis, and multiple organ failure are associated with critically ill COVID-19 patients, consequently resulting in fatalities (5–7).

Coronaviruses (CoVs) belong to the family Coronaviridae, and all species responsible for severe acute respiratory syndrome (SARS) fall under the genus beta-coronavirus (8), most of which is enzootic, and only a few are known to infect humans directly. Having established an animal-human host transmission, the lethality of CoVs has been demonstrated by past outbreaks of SARS and the Middle East respiratory syndrome (MERS) in the years 2003 and 2012, resp. (9–11).

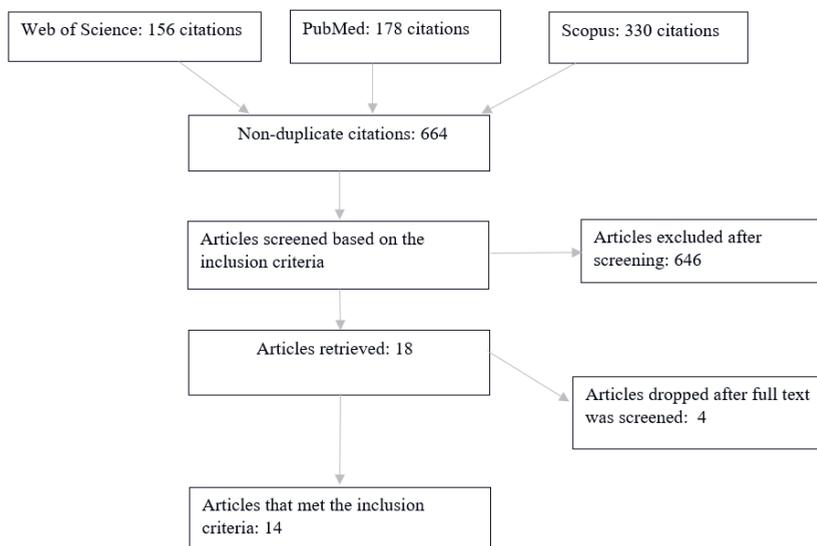
The whole viral genome of the novel SARS-CoV-2 has a 96 % similarity to the bat CoV and 79.6 % sequence identity to SARS-CoV (12). Empirical evidence has shown that although SARS-CoV-2 (reported as 2019-nCoV) is closer to bat-SL-CoVZXC21 and bat-SL-CoVZC45 at the whole-genome level, the receptor-binding domain of SARS-CoV-2 falls within a lineage closer to that of SARS-CoV (13). Genome transcription of beta-coronaviruses yields a polypeptide of approximately 800 kDa, which produces several proteins upon proteolytic cleavage, a process mediated by either the papain-like protease (PLpro) or 3-chymotrypsin-like protease (3CLpro) (14, 15). In addition, SARS-CoV emerged from a zoonotic reservoir and coupled with cytokine, chemokine, and interferon-stimulated gene (ISG) responses in patients, evidence that SARS-CoV pathogenesis is partially controlled by innate immune signaling (16–19). The drug targets among coronaviruses include the main protease (Mpro also called 3CLpro) and papain-like protease(s). The Mpro is liable to block viral replication, meanwhile, papain-like protease (PLpro) is essential for processing the polyproteins translated from the viral RNA (14, 21–23). Thus, 3CLpro and PLpro are validated drug targets for developing antiviral agents against CoVs.

Although the reported cases of previous SARS outbreaks were confined in Asia, the magnitude of the COVID-19 pandemic has presented a more insidious threat to global health and man's livelihood. Nonetheless, the identification of new drugs with high efficacy against CoVs is still elusive. As the search for drugs to combat COVID-19 continues, plant-derived compounds present a catalogue of potential anti-SARS-CoV-2 therapeutics, recording significant inhibitory effects on SARS-associated CoVs (24–26). These natural products provide active pharmaceutical ingredients and structural blueprints for designing their synthetic analogues with improved antiviral activity (27). Therefore, the present review aims to systematically evaluate existing reports on the anti-SARS-CoV activities of medicinal plants and their associated bioactive compounds to identify potential drug candidates for COVID-19 therapy.

## METHODOLOGY AND DATA SOURCES

### *Data curation*

The articles subjected to meta-analysis were extracted from the following databases: PubMed, Web of Science and Scopus. These databases were searched within English language papers published between 2005 and 2020, on medicinal plants used in the treatment of SARS-COV infection. The databases were searched using a combination of the following keywords: “coronavirus”, “SARS-CoV”, “COVID-19”, “medicinal plants”, “traditional medicine”, “Chinese medicine”, “plant extract”, “cysteine protease”, “severe acute respiratory syndrome”, “SARS-CoV-1 or SARS-COV-1”, “SARS-CoV-2 or SARS-COV-2”, “herbs”, “SARS-CoV 3CLpro”, “SARS-CoV PLpro” and “antiviral agent”. A total of 664 published articles between the years 2005 and 2020 were identified; a schematic representation of the selection process for reviewed articles is given in Scheme 1. As a complementary procedure, the relevant studies were checked manually for any citation missed by the electronic database.



Scheme 1

### *Characteristic evaluation and inclusion barometer*

The systematic review was achieved using the PRISMA guidelines protocol (28). Eligibility criteria were set as follows: articles written in English, articles published between years 2005 and 2020, medicinal plants tested against SARS-CoV 3CLpro and PLpro enzymes, and their respective isolated compounds. The exclusion criteria include animal and clinical trial studies. Notably, studies devoid of either mean or standard deviation of inhibitory potencies were also excluded from the meta-analysis to maintain the quality of the findings.

### *Data synthesis and statistical analysis*

The retrieved data were statistically analyzed, and the Stata 15.0 (Stata Corp, College Station, TX, USA) was used for the graphical representation of the pooled data. Statistical heterogeneity was assessed by both a Cochran's chi-squared test ( $Q$  test) and an  $I^2$ -squared test. A fixed-effects model was used when there was no significant statistical heterogeneity ( $p > 0.1$  and an  $I^2$  value  $< 50\%$ ). In other cases, a random effects model statistical approach was employed. In this study, because the extracted articles were from the general population, a random effects meta-analysis was considered to be taken from an inverse-variance model. Effect sizes (ES) were estimated using the forest plot as a prelude for heterogeneity and biases examinations. In this study, the random effects model was applied to estimate and detect sources of statistical heterogeneity that may arise for different reasons. Furthermore, subgroup analysis was conducted to test whether there are subsets of the included studies that capture the pooled ES. The funnel plot and Egger's tests were simultaneously used to assess potential publication biases.

## RESEARCH OUTCOMES

### *Meta-analysis*

A forest plot is an orthodox device used to visualize how the estimate of ES of each study is distributed around a zero or pooled effect estimate. The ES estimate of each study is represented in the forest plot as a square box. The area of each box represents the weight of each study contributing to the pooled estimate while the center of a diamond equals the pooled effect estimate. The ends of the diamond indicate the limits of 95 % confidence interval (CI). Hence, the heterogeneity test and  $Q$  statistics gave significantly large value (chi-square = 5860.22,  $df = 19$ ,  $p < 0.0001$ ,  $I^2 = 99.7\%$ ), indicating the presence of enormous variation among studies. The residual amount of heterogeneity indicates the extent of variability as compared with the effect size. Besides, the percentage of total variation resulting from heterogeneity across studies is substantial for  $I^2$ . These findings generally imply that the proportion of total variance among pooled studies (*i.e.*,  $IC_{50}$  of the active compounds) can be attributed to the accuracy of evaluation of heterogeneity in the effect sizes. The pooled estimated mean using the fixed effect model showed significant heterogeneity between the studies. Hence, we performed the analyses using the random effects model. Using the random-effects model, the estimated pooled mean of potential anti-SARS-CoV compounds based on the  $IC_{50}$  was 6.12 % (95 % CI 6.09-6.16) with significant heterogeneity between studies ( $I^2 = 99.7\%$ ,  $p < 0.0001$ ). The pooled estimated mean of the potential anti-SARS-CoV compounds based on the  $IC_{50}$  is presented using a forest plot (Fig. 1).

The main void of the heterogeneity concept is that it provides only global measures without additional information about the sources of heterogeneity. The inherent void demands that subgroup analysis is to be performed to unveil the sources of heterogeneity. Subgroup analysis is the splitting of the participant data into subgroups to establish comparisons between sub-data. The interpretation of subgroup meta-analysis can lead to informative insights into the proper implication that would not be obtained from the non-subgroup analysis. Thus, an analysis of the isolated compounds subgroup was conducted to assess the potential heterogeneity between the studies included in the meta-analysis.

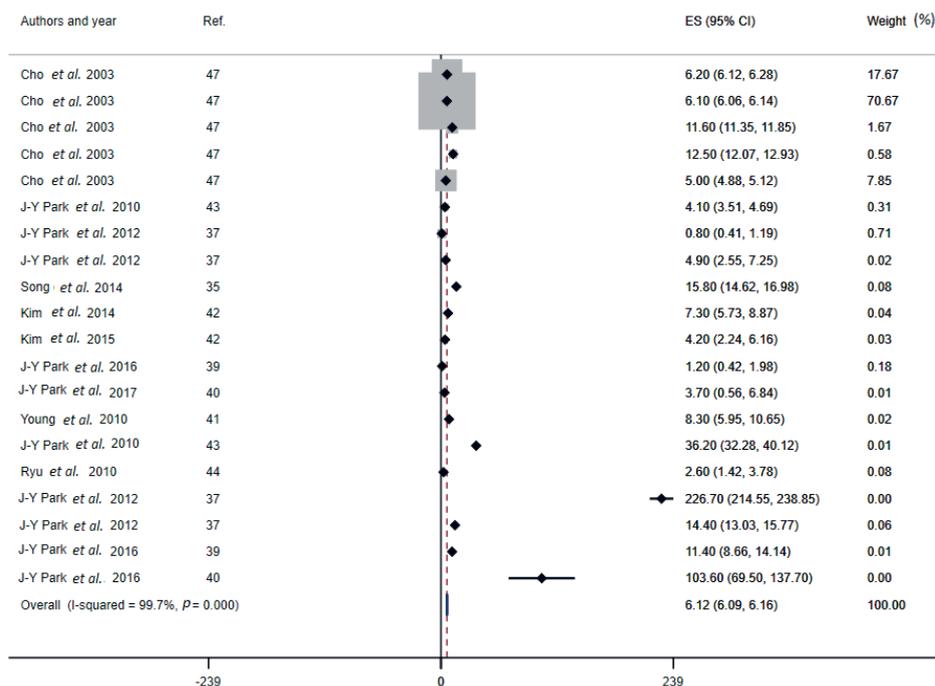


Fig. 1. Overall pooled mean estimate obtained for active chemical constituents ( $IC_{50}$ ) tested against SARS-CoV proteases.

Further, the subgroup analysis was conducted using 3CLpro and PLpro inhibitors to assess the potential heterogeneity between the studies; the findings established a statistically significant difference ( $p < 0.001$ ) in the subgroup. Of the 20 studies, the highest pooled estimated mean was found in studies reporting 3CLpro inhibitors [10.07 % (95 % CI: 9.29–10.85),  $I^2 = 99.7$  %], followed by the studies conducted with PLpro inhibitors [6.12 % (95 % CI: 6.08–6.15),  $I^2 = 99.7$  %] (Fig. 2). This result suggests that the inhibitory potencies ( $IC_{50}$ ) against 3CLpro and PLpro were significantly different among the active compounds.

Furthermore, one of the medicinal plants active against SARS-CoV *in vitro* includes *Tribulus terrestris*. The plant belongs to the genus *Tribulus* (Zygophyllaceae), a large, heterogeneous and widely dispersed genus comprising of twenty-seven species (29, 30). *T. terrestris*, the most researched species, is rich in steroids, saponins, flavonoids, sterols, lignan amides and cinnamic acid (30–32). The pharmacological applications of *T. terrestris* such as anticancer, antioxidant, anti-inflammatory, antidiuretic, and antimicrobial have been reported (33, 34). Song *et al.* (35) revealed that the methanolic extracts of *T. terrestris* fruits showed superior inhibitory activity towards SARS-CoV PLpro compared to ethyl acetate, hydroalcoholic and aqueous extracts. Purification of the isolated compounds from the methanolic extracts unveiled cinnamic amide derivatives. All the isolated compounds displayed significant PLpro inhibition with  $IC_{50}$  values of 15.8–70.1  $\mu\text{mol L}^{-1}$  (Table I). The highest inhibitory potency was observed for terrestrimine (1) and terrestramide (2) with  $IC_{50}$  of  $15.8 \pm 0.6$  and  $21.5 \pm 0.5 \mu\text{mol L}^{-1}$ , resp. (Fig. 3).

Table 1. Plant extracts and isolated compounds with promising SARS-CoV inhibitory potency

| Plant (genus/species)                       | Family          | Extraction medium                           | SARS-CoV-1 protease tested | Plant part studied | Isolated compound(s)  | Remark(s)   | Ref. |
|---|-----------------|---|----------------------------|--------------------|---|---|------|
| <i>Tribulus terrestris</i> L.               | Zygophyllaceae  | Ethyl acetate, methanol, aq. ethanol, water | PLpro                      | Fruits             | From methanolic extract: <i>N-trans</i> -caffeoyltyramine, <i>N-trans</i> -coumaroyl-tyramine, <i>N-trans</i> -feruloyl-tyramine, terrestrinamide (1), <i>N-trans</i> -feruloyl-octopamine, terrestrimine (2) | All extracts (excluding aq. extracts) show good inhibitory potency against PLpro at 300 µg mL <sup>-1</sup>   | 35   |
| <i>Isatis indigotica</i> L.                 | Brassicaceae    | Water                                       | 3CLpro                     | Root               | Indigo, indirubin, indican, sinigrin (3), beta-sitosterol, aloemodin, hesperetin (4), quercetin, naringenin, daidzein, emodin, chrysohanol  | 50 % inhibitory conc. of aq. extracts: 191.6 ± 8.2 µg mL <sup>-1</sup> for the cell-based cleavage, 53.8 ± 4.2 µg mL <sup>-1</sup> for cell-free cleavage assay | 36   |
| <i>Salvia miltiorrhiza</i> Bunge            | Lamiaceae       | Ethanol                                     | 3CLpro, PLpro              | Root               | Tanshinone IIA (9), tanshinone IIB (11), methyl tanshinonate (8), cryptotanshinone (6), tanshinone I (9), dihydrotanshinone I (5), rosmariquinone (10)  | Ethanol extracts inhibit 60 and 88 % 3CLpro and PLpro at 30 µg mL <sup>-1</sup> , resp.   | 37   |
| <i>Taxus celebica</i> (Warb) H.L.Li         | Taxaceae        | Ethanol                                     | 3CLpro                     | Not reported       | Not reported  | 12 %  | 38   |
| <i>Uvaria macrocarpa</i> (Champ. ex Benth.) | Annonaceae      | Ethanol                                     | 3CLpro                     | Not reported       | Not reported  | 21.4 %  | 38   |
| <i>Rubus suaveolens</i> S. Lee              | Rosaceae        | Water                                       | 3CLpro                     | Not reported       | Not reported  | 0 %   | 38   |
| <i>Auricularia auricular</i> (L.) Underw    | Auriculariaceae | Water                                       | 3CLpro                     | Not reported       | Not reported  | Inhibition rate by the respective extract at 100 µg mL <sup>-1</sup>  | 38   |
| <i>Glycyrrhiza uralensis</i> Fisch. ex DC.  | Leguminosae     | Water                                       | 3CLpro                     | Root               | Not reported  | 13.5 %  | 38   |
| <i>Mangifera indica</i> L.                  | Anacardiaceae   | Water                                       | 3CLpro                     | Leaves             | Not reported  | 3.5 %   | 38   |
| <i>Scutellaria baicalensis</i> Georgi       | Lamiaceae       | Water                                       | 3CLpro                     | Leaves             | Not reported  | 13.6 %  | 38   |
| <i>Sophora flavescens</i> Aiton             | Fabaceae        | Ethanol                                     | 3CLpro                     | Radix              | Not reported  | 20.2 %  | 38   |

| Plant (genus/species)                          | Family          | Extraction medium | SARS-CoV-1 protease tested | Plant part studied | Isolated compound(s)  | Remark(s)  | Ref. |
|--|-----------------|-------------------|----------------------------|--------------------|---|--|------|
| <i>Cyrtomium fortunei</i> J.Sm.                | Dryopteridaceae | Ethanol           | 3CLpro                     | Rhizome            | Not reported  | 25.7 %   | 38   |
| <i>Brucea javanica</i> (L.) Merr.              | Simaroubaceae   | Ethanol           | 3CLpro                     | Fruits             | Not reported  | 5.3 %  | 38   |
| <i>Rheum palmatum</i> L.                       | Polygonaceae    | Ethanol           | 3CLpro                     | Herbs              | Extracts/fractions: RH10, RH11, RH12, RH12L, RH122, RH124, RH125  | Chloroform-methanol extracts markedly inhibit 3C-like protease; RH12L fraction the most active ( $IC_{50} = 13.76 \pm 0.03 \mu\text{g mL}^{-1}$ ), with inhibition rate up to 96 % | 38   |
| <i>Angelica keiskei</i> Ito                    | Apiaceae        | Ethanol           | 3CLpro, PLpro              | Leaves             | Isobavachalcone (12), 4-hydroxyderricin (13), xanthoangelol (14), xanthoangelol B, D, E, G, F (15–19), xanthokeistal A (20), psoralen (21), bergapten (22), xanthotoxin (23), isopimpinellin (24)   | The ethanolic extract shows 75 % inhibition of 3CLpro and 88 % inhibition of PLpro at 30 $\mu\text{g mL}^{-1}$ , resp.   | 39   |
| <i>Broussonetia papyrifera</i> (L.) Vent.      | Moraceae        | Ethanol           | 3CLpro, PLpro              | Root               | Broussonetolone A, B; 4-hydroxy-isolonchocarpin, papyriflavonol A (25), 3-(3-methylbut-2-enyl)-3',4',7-trihydroxyflavane, kazinol A, B, F, J; broussonetolone A   | Papyriflavonol A most potent PLpro inhibitor ( $IC_{50} = 3.71.6 \mu\text{mol L}^{-1}$ ), but poor inhibitor for 3CLpro ( $IC_{50} = 103.6 \pm 17.4 \mu\text{mol L}^{-1}$ )        | 40   |
| <i>Torreya nucifera</i> (L.) Siebold and Zucc. | Taxaceae        | Ethanol           | 3CLpro                     | Leaves             | Amentoflavone (34), 18-hydroxyferruginol (26), hinokiol (27), ferruginol (28), 18-oxoferruginol, O-acetyl-18-hydroxyferruginol (29), methyl dehydroabietate (31), bilobetin (35), ginkgetin (36), isopimaric acid (32), sciadopitysin (37), kayadiol (33) | Ethanol extract exhibits good 3CLpro inhibitory activity: 62 % at 100 $\mu\text{g mL}^{-1}$  | 41   |
| <i>Psoralea corylifolia</i> L.                 | Fabaceae        | Ethanol           | PLpro                      | Seeds              | Bavachinin, neobavaisoflavone, isobavachalcone (41), 4'-O-methyl-bavachalcone, psoralidin (42), corylifol A   | Ethanol extract shows 50 % inhibition at 15 $\mu\text{g mL}^{-1}$  | 42   |

| Plant (genus/species)                                     | Family        | Extraction medium                | SARS-CoV-1 protease tested | Plant part studied     | Isolated compound(s)  | Remark(s)  | Ref. |
|---|---------------|----------------------------------|----------------------------|------------------------|---|--|------|
| <i>Alnus japonica</i> (Thunb.) Steud.                     | Betulaceae    | Ethanol                          | 3CLpro, PLpro              | Stem bark              | Platyphyllene, hirsutenone (43), platyphyllone, platyphyllanol-5-xylopyranoside, hirsutanol, oregonin, rubranol, rubranoside B and rubranoside A  | Hirsutenone most potent PLpro inhibitor ( $IC_{50} = 4.1 \mu\text{mol L}^{-1}$ ) | 43   |
| <i>Tripterigium regelii</i> (Sprag & Takeda)              | Celastraceae  | Methanol                         | PLpro                      | Bark                   | Celastrin (46), pristimerin (47), tingenone (45), iguesterin (44)   | MeOH (95 %) extracts inhibit >70 % 3CLpro activity at $30 \mu\text{g mL}^{-1}$   | 44   |
| <i>Camellia sinensis</i> var. <i>sinensis</i>             | Theaceae      | Water                            | 3CLpro                     | Green tea              | Caffeine, theophylline, catechin (C), epigallocatechin, (-)-epigallocatechin gallate, epicatechin, epicatechin gallate  | $125 \mu\text{g mL}^{-1}$  | 45   |
| <i>Camellia sinensis</i> (L.) Kuntze var. <i>assamica</i> | Theaceae      | Water                            | 3CLpro                     | Black tea              | Theaflavin (TF1), mixture of theaflavin-3'-gallate (TF2b) and theaflavin-3-gallate (TF2a), theaflavin-3,3'-digallate (TF3) (48), isotheaflavin-3'-gallate (TF2B) (49), tannic acid (50) | 70 $\mu\text{g mL}^{-1}$<br>50 % inhibition                                      | 45   |
| <i>Camellia sinensis</i> (L.) Kuntze var. <i>assamica</i> | Theaceae      | Water                            | 3CLpro                     | Oolong tea, Pu-erh tea | Caffeine, theophylline, catechin (C), epigallocatechin, (-)-epigallocatechin gallate, epicatechin, epicatechin gallate isolated from oolong tea<br>Not reported for Pu-erh tea          | $125$ and $25 \mu\text{g mL}^{-1}$ , resp.                                       | 45   |
| <i>Gentiana scabra</i> Bunge                              | Gentianaceae  | Ethanol, water, methanol, hexane | 3CLpro                     | Rhizome                | Not reported  | Hexane extracts at $> 50 \mu\text{g mL}^{-1}$                                    | 46   |
| <i>Dioscorea batatas</i> Decne                            | Dioscoreaceae | Ethanol, water, methanol, hexane | 3CLpro                     | Tuber                  | Not reported  | 50 % inhibition by the respective extract  | 46   |
| <i>Cassia tora</i> (L.) Roxb                              | Leguminosae   | Ethanol, water, methanol, hexane | 3CLpro                     | Seed                   | Not reported  | Hexane extracts at $> 50 \mu\text{g mL}^{-1}$                                    | 46   |

| Plant (genus/species)  | Family        | Extraction medium                | SARS-CoV-1 protease tested | Plant part studied | Isolated compound(s)  | Remark(s)  | Ref. |
|--|---------------|----------------------------------|----------------------------|--------------------|---|--|------|
| <i>Taxillus chinensis</i> (DC.) Danser                             | Loranthaceae  | Ethanol, water, methanol, hexane | 3CLpro                     | Stem, leaves       | Not reported  | Hexane extracts at > 50 µg mL <sup>-1</sup>  | 46   |
| <i>Cibotium barometz</i> (L.) J.Sm.                                | Cibotiaceae   | Ethanol, water, methanol, hexane | 3CLpro                     | Rhizome            | Not reported  | Ethanol and methanol extracts at > 50 and 39 µg mL <sup>-1</sup> , resp.   | 46   |
| <i>Paulownia tomentosa</i> (Thunb.) Steud.                         | Paulowniaceae | Methanol                         | PLpro                      | Fruit              | Tomentin A-E (51-55), 3'-O-methyl-diplacol (56), 4'-O-methyl-diplacol (60), 3'-O-methyl-diplacone (57), 4'-O-methyl-diplacone (58), mimulone (59), diplacone (61), 6-geranyl-4',5,7-trihydroxy-3',5'-dimethoxy-flavanone (62) | Tomentin A-E show excellent inhibitory activity (IC <sub>50</sub> from 5 to 13 µmol L <sup>-1</sup> )  | 47   |
| <i>Chamaecyparis obtuse</i> (Siebol. & Zucc) var. <i>formosana</i> | Cupressaceae  | Ethyl acetate                    | 3CLpro                     | Heart-wood         | Ferruginol (63), dehydroabieta-7-one (64), sugiol (66) 8-hydroxy-abieta-9(11),13-dien-12-one (67), 6,7-dehydroroyleanone (69), pinusolidic acid (71), R-cadinol (74), hinokinin (77), savinin (78)                            | Betulinic acid (not isolated) and savinin competitive inhibitors of SARS-CoV 3CL protease (IC <sub>50</sub> = 10 and 25 µmol L <sup>-1</sup> , resp) | 25   |
| <i>Juniperus formosana</i> Hayata                                  | Cupressaceae  | Ethyl acetate                    | 3CLpro                     | Heart-wood         | 3β,12-diacetoxyabieta-6,8,11,13-tetraene (70), cedrane-3a,12-diol (73), betulonic acid (76), cryptojaponol (66)   | Betulonic acid inhibits 3CL protease 0.63 µmol L <sup>-1</sup> at 50 %   | 25   |
| <i>Cryptomeria japonica</i> (L.f.) D.Don                           | Cupressaceae  | Ethyl acetate                    | 3CLpro                     | Heart-wood         | 7β-hydroxydeoxycryptojaponol  | 7β-hydroxydeoxycryptojaponol inhibits 3CL protease 1.15 µmol L <sup>-1</sup> at 50 %   | 25   |

The roots of *Isatis indigotica*, a member of the family Cruciferae, native to China, are known for their potency against influenza, hepatitis A and Japanese encephalitis (48–50). Lin *et al.* (36) examined the efficacy of five major compounds of *I. indigotica* root, namely, indigo, indirubin, indican, sinigrin and beta-sitosterol, and seven plant-derived phenolic compounds, namely, aloe emodin, hesperetin, quercetin, naringenin, daidzein, emodin and chrysophanol, which were tested for anti-SARS-CoV 3CLpro effects using cell-free and cell-based cleavage assays. Only two phenolic compounds, aloe emodin and hesperetin, isolated from the aqueous extracts of the plant's root, dose-dependently inhibited cleavage activity of the 3CLpro, in which the  $IC_{50}$  was 366  $\mu\text{mol L}^{-1}$  for aloe emodin and 8.3  $\mu\text{mol L}^{-1}$  for hesperetin in the cell-based assay (Table I). Sinigrin (3) ( $IC_{50} = 217 \mu\text{mol L}^{-1}$ ) and hesperetin (4) ( $IC_{50} = 8.3 \mu\text{mol L}^{-1}$ ) shown in Fig. 4 emerged as potential leads for developing inhibitors of SARS-CoV 3CLpro. Sinigrin was more efficient in blocking the cleavage processing of 3CLpro than indigo ( $IC_{50} = 300 \mu\text{mol L}^{-1}$ ). The report from the literature accredited the antiviral effect of sinigrin (3) and hesperetin (4), including poliovirus, pseudorabies virus, Sindbis virus, herpes simplex virus types 1 and 2, parainfluenza virus and vaccinia virus (51–53). In addition, the most potent compounds, *i.e.*, sinigrin (3) and hesperetin (4) with  $CC_{50}$  of above 2  $\text{mmol L}^{-1}$  were considerably less cytotoxic to Vero cells than

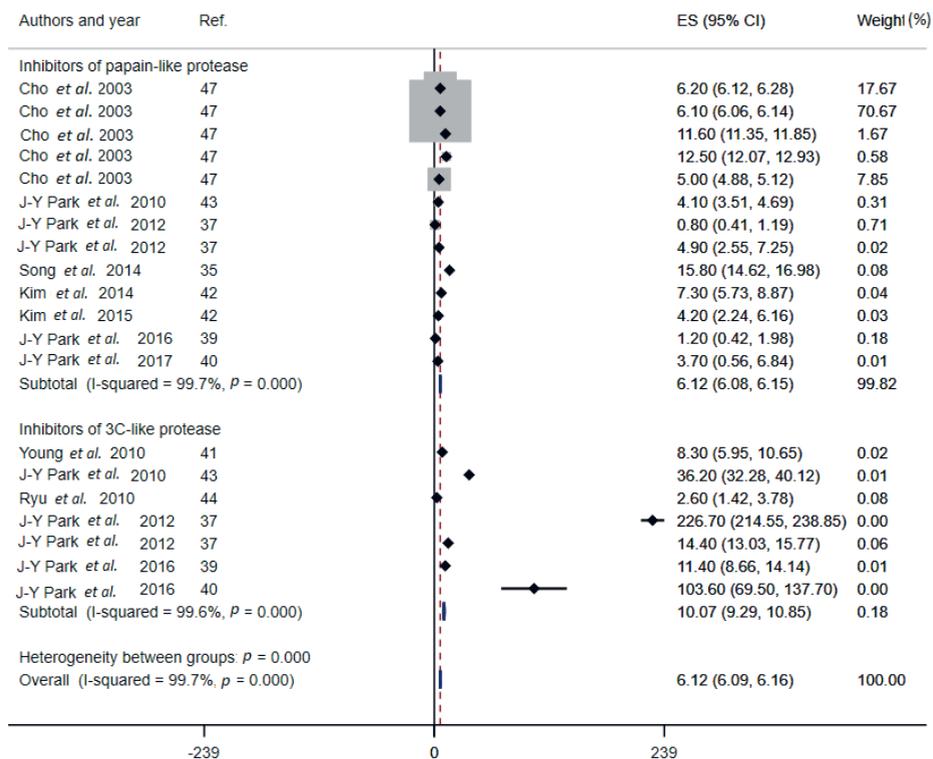


Fig. 2. Sub-group analysis of the active chemical constituents ( $IC_{50}$ ) tested against 3CLpro and PLpro SARS-CoV.

indigo and beta-sitosterol. Therefore, they may be considered as potential leads in the development of inhibitors of SARS-CoV and SARS-CoV-2 3CLpro.

Similarly, in a quest to find inhibitors of viral replication in SARS-CoV, Ji-Y Park *et al.* (37) focused on the inhibitory action of naturally derived tanshinones against 3CLpro and PLpro of the virus. All the isolates were found in the lipophilic fraction (*n*-hexane) of *S. miltiorrhiza* extracts. The 3CLpro inhibitory potency of the compounds, dihydrotanshinone I (5), cryptotanshinone (6), tanshinone IIB (7), methyl tanshinonate (8), tanshinone I (9), rosmariquinone (10) and tanshinone IIA (11), ranged from 14.4 to 226.7  $\mu\text{mol L}^{-1}$ , whereas all the isolated compounds (5–11) showed inhibitory activities to both 3CLpro and PLpro. The activity was significantly affected by subtle changes in the structure. Notably, dihydrotanshinone I (5) ( $IC_{50} = 14.4 \mu\text{mol L}^{-1}$ ) showed ~16-fold superior potency compared to cryptotanshinone (6) ( $IC_{50} = 226.7 \mu\text{mol L}^{-1}$ ) (Fig. 5). Cryptotanshinone (6) exhibited the lowest inhibitory activity compared to the compounds with furan moiety. The introduction of the hydroxy-methyl group on the D-ring of tanshinone IIB (7) increased its enzyme inhibitory activity with an  $IC_{50}$  value of 24.8  $\mu\text{mol L}^{-1}$  (Fig. 5). The corresponding methyl ester on the D-ring, methyl tanshinonate (8), also showed a similar potency enhancement ( $IC_{50} = 21.1 \mu\text{mol L}^{-1}$ ). In contrast, the dihydrofuran moiety of dihydrotanshinone ( $IC_{50} = 14.4 \mu\text{mol L}^{-1}$ ) showed higher inhibitory activity against 3CLpro than tanshinone I (9) ( $IC_{50} = 38.7 \mu\text{mol L}^{-1}$ ).

The isolated compounds were also tested against PLpro, and surprisingly, cryptotanshinone (6) displayed the most potent inhibitory activity ( $IC_{50} = 0.8 \mu\text{mol L}^{-1}$ ), whereas tanshinone I (9) ( $IC_{50} = 8.8 \mu\text{mol L}^{-1}$ ) and dihydrotanshinone I (5) ( $IC_{50} = 4.9 \mu\text{mol L}^{-1}$ ) exhibited similar inhibitory potencies possibly due to their identical ring-A structure (Fig. 5).

Interestingly, the structurally related abietane analog, rosmariquinone (10), displayed significant activity against both 3CLpro and PLpro with  $IC_{50}$  values of 21.1 and 30.0

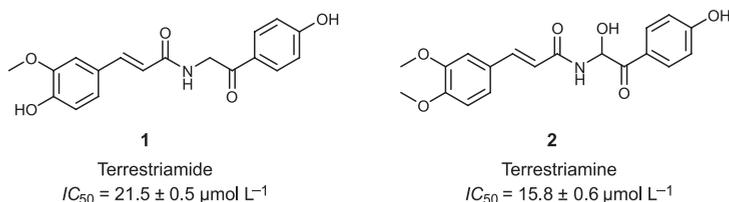


Fig. 3. Chemical structure of terrestriamide and terrestriamine isolated from *T. terrestris* fruits.

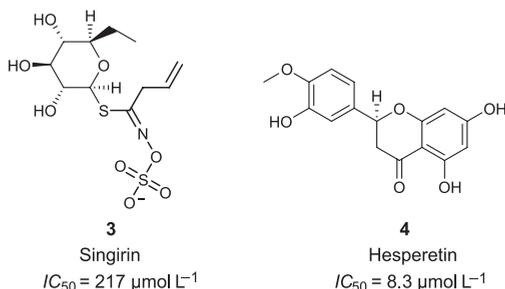
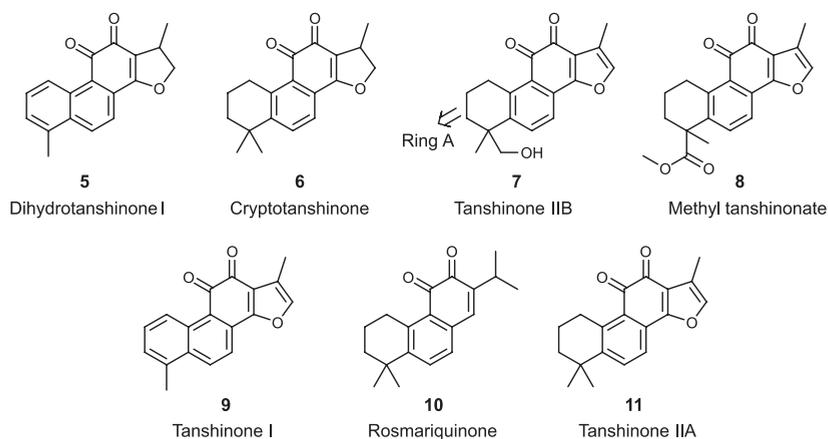


Fig. 4. Chemical structure of singirin and hesperetin isolated from *I. indigotica*.

$\mu\text{mol L}^{-1}$ , resp. The introduction of a three-ringed abietane analog, rosmariquinone (**10**), showed simple reversible slow-binding inhibitor and mixed-type inhibition. In addition, tanshinone I (**9**) showed the most potent DUB activity with an  $IC_{50}$  value of  $0.7 \mu\text{mol L}^{-1}$ . The results from the isolated compounds merit further examination for their effect on the inhibition of SARS-CoV-2.

Luo *et al.* (38) reported twelve plants as 3CLpro inhibitors (Table I). The extracts/fractions of *Rheum palmatum* L. such as RH10, RH11, RH12, RH121, RH122, RH124 and RH125 significantly inhibited SARS coronavirus 3C-like protease. Fraction RH121 ( $IC_{50} = 13.76 \pm 0.03 \mu\text{g mL}^{-1}$ ) (Fig. 6) emerged as highly potent anti-SARS-CoV therapeutic agent. The ethanolic extracts of rhubarb showed no cytotoxicity at  $20 \text{ mg mL}^{-1}$ , which partly makes them an excellent tool for anti-coronavirus drug screening. Rhubarb is plentiful in China as traditional medicine for viral diseases.

Ji-Y Park *et al.* (39) showed the significant inhibition of 3CLpro and PLpro by EtOAc-soluble fraction of ethanolic extract of *Angelica keiskei* (75 and 88 % inhibition at  $30 \text{ mg mL}^{-1}$ , resp.). Nine alkylated chalcones, namely isobavachalcone (**12**), 4-hydroxyderricin



| Compd.              | $IC_{50}$ ( $\mu\text{mol L}^{-1}$ ) |                |
|---------------------|--------------------------------------|----------------|
|                     | 3CLpro                               | PLpro          |
| Tanshinone IIA      | $89.1 \pm 5.2$                       | $1.6 \pm 0.5$  |
| Tanshinone IIB      | $24.8 \pm 5.2$                       | $10.7 \pm 1.7$ |
| Methyl tanshinonate | $21.1 \pm 0.8$                       | $9.2 \pm 2.8$  |
| Cryptotanshinone    | $226.7 \pm 6.2$                      | $0.8 \pm 0.2$  |
| Tanshinone I        | $38.7 \pm 8.2$                       | $8.8 \pm 0.4$  |
| Dihydrotanshinone I | $14.4 \pm 0.7$                       | $4.9 \pm 1.2$  |
| Rosmariquinone      | $21.1 \pm 0.8$                       | $30.0 \pm 5.5$ |

Fig. 5. Chemical structures and inhibitory effects of isolated compounds from *S. miltiorrhiza*.

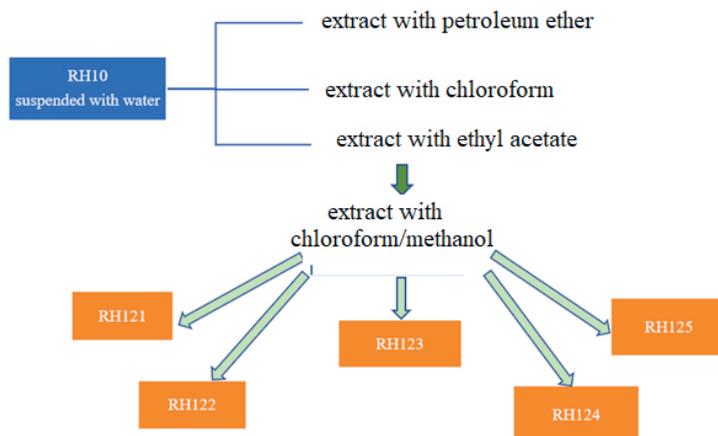


Fig. 6. Flow diagram of separation of various extracts/fractions from *Rheum palmatum* L. using different solvents.

(13), xanthoangelol (14), xanthoangelol F (15), xanthoangelol D (16), xanthoangelol E (17), xanthoangelol B (18), xanthoangelol G (19), xanthokeistal A (20), and psoralen (21) and four coumarins, bergapten (22), xanthotoxin (23) and isopimpinellin (24) were isolated from *Angelica keiskei*, and the inhibitory activities of these constituents against SARS-CoV 3CLpro and PLpro were reported. Among the different chalcones, flavanones, and coumarins isolated from the plant (Table I), xanthoangelol E (12) containing the perhydroxyl group showed 3CLpro and PLpro inhibitory potencies ( $IC_{50} = 11.4$  and  $1.2 \mu\text{mol L}^{-1}$ ) that are 5- to 40-fold superior to other analogs (Fig. 7). The structure-activity relationship analysis showed that the peroxide unit on hemiterpene might influence the polyhydroxylated chain's binding and conformational stability through intramolecular hydrogen bonding. The optimization of this compound in the development of protease inhibitors may yield an effective anti-SARS-CoV-2 agent.

Papyriflavonol A (24), a polyphenol, has been reported as the most potent PLpro inhibitor with an  $IC_{50}$  value of  $3.7 \mu\text{mol L}^{-1}$  compared to other isolated polyphenols from *Broussonetia papyrifera* (40). All the isolated polyphenols were more potent against PLpro than 3CLpro (Table I). An evaluation of the structure-activity relationship revealed that the prenyl groups' position was beneficial to inhibitory potency of papyriflavonol A (24) (Fig. 8). Hence, the significant activity of this compound showed that it can be further developed as an anti-coronavirus agent targeting PLpro and 3CLpro proteases.

Ryu *et al.* (41) reported the SARS-CoV 3CL pro-activity of eight diterpenoids and four bioflavonoids isolated from *Torreya nucifera*. *T. nucifera* is a slow-growing, coniferous tree native to snowy areas near the Sea of Jeju Island in Korea (54). The traditional use of the plant in Asian medicine as a remedy for stomachache, hemorrhoids, and rheumatoid arthritis was reported by Bae *et al.* (55). The pharmacological activity of *T. nucifera* also included antioxidative, antiproliferative, anti-inflammatory, hepatoprotective, and neuroprotective ones (56–58). The isolated diterpenes, namely, 18-hydroxyferruginol (26), hinokiol (27), ferruginol (28), 18-oxoferruginol (29), *O*-acetyl-18-hydroxyferruginol (30),

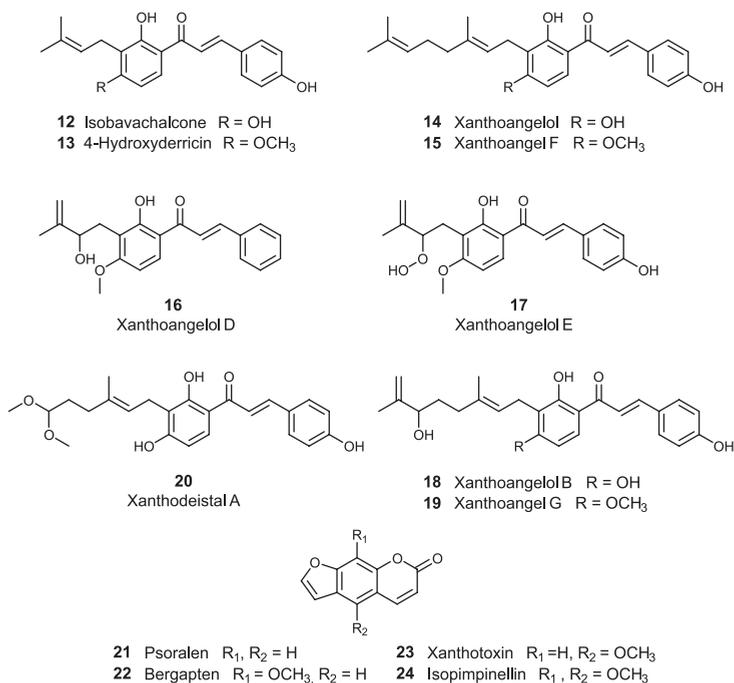


Fig. 7. Chemical structures of chalcones and coumarins isolated from *A. keiskei*.

methyl dehydroabietate (31), isopimaric acid (32) and kayadiol (33), and bioflavonoids amentoflavone (34), bilobetin (35), ginkgetin (36) and sciadopitysin (37) from *T. nucifera* (41) were tested in parallel with the standard flavones apigenin (38), luteolin (39) and quercetin (40) (Fig. 9). The latter compounds were included to establish the structure-activity relationship of biflavones and they inhibited 3CLpro activity with  $IC_{50}$  values of 280.8, 20.2, and 23.8  $\mu\text{mol L}^{-1}$ , resp.

The eight in-house diterpenoid libraries tested against SARS-CoV 3CLpro showed that ferruginol (28) exhibited superior inhibitory activity ( $IC_{50} = 49.6 \mu\text{mol L}^{-1}$ ) which was approximately four-fold more potent than that of abietic acid ( $IC_{50} = 189.1 \mu\text{mol L}^{-1}$ ). Intro-

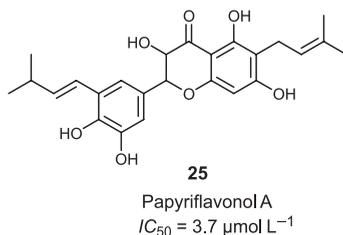


Fig. 8. Chemical structure of papyriflavonol A, the promising anti-SARS-CoV compound, isolated from *B. papyrifera*.

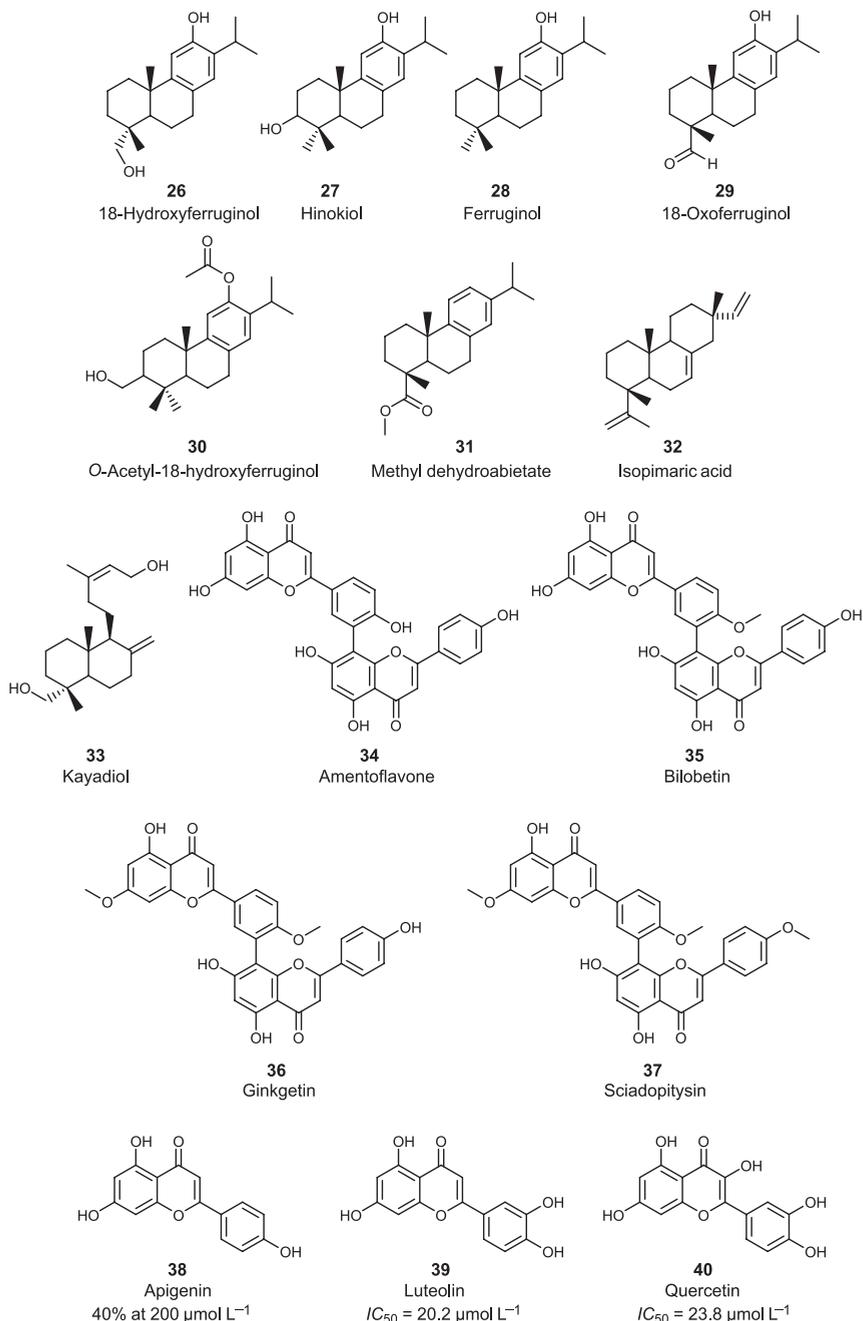


Fig. 9. Chemical structures of compounds isolated from the leaves of *T. nucifera* tested against SARS-CoV 3CLpro.

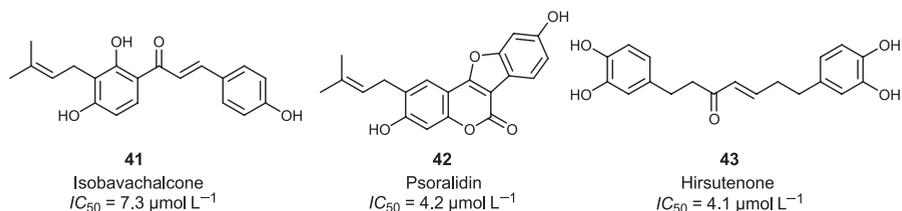


Fig. 10. Chemical structures of some promising compounds (isobavachalcone, psoralidin, hirsutenone) showing anti-SARS-CoV properties, isolated from *Alnus japonica*.

duction of methoxy group to amentoflavone (**34**) moiety to give bilobetin (**35**), ginkgetin (**36**) and sciadopitysin (**37**) bioflavonoids, resulted in the less potent inhibitory activity of these compounds ( $IC_{50} = 32.0\text{--}72.3 \mu\text{mol L}^{-1}$ ). The methoxy group at position C-7 of ginkgetin (**36**) with  $IC_{50}$  of  $32.0 \mu\text{mol L}^{-1}$  and sciadopitysin (**37**) with  $IC_{50}$  of  $38.4 \mu\text{mol L}^{-1}$  might be responsible for a two-fold increase in the anti-SARS-CoV 3CLpro inhibitory activity compared to bilobetin (**35**), with hydroxyl functional group at position C-7 ( $IC_{50} = 72.3 \mu\text{mol L}^{-1}$ ). The most potent inhibitor, amentoflavone (**34**) exhibited an  $IC_{50}$  value of  $8.3 \mu\text{mol L}^{-1}$  toward SARS-CoV 3CLpro, making this compound about 30-fold more potent than apigenin. Meanwhile, the inhibitory activity of luteolin (**39**) ( $IC_{50} = 20.2 \mu\text{mol L}^{-1}$ ) was inferior to amentoflavone (**34**) inhibitory activity. The apigenin motif in amentoflavone has possibly played a pivotal role in the SARS-CoV 3CLpro inhibition.

*Psoralea corylifolia* (Leguminosae) is used as a food additive and is distributed from India to Southeast Asia. The seeds are found to be helpful as a tonic or an aphrodisiac (50, 59). Moreover, the phytochemicals from *Psoralea corylifolia* demonstrate a wide range of biological activities such as antioxidant, antibacterial, anti-inflammatory, antidepressant, and antiviral (60–69). Kim and coworkers (42) have also shown that the ethanolic extracts of *P. corylifolia* seeds exhibit potent inhibitory potency against SARS-CoV PLpro. Fraction

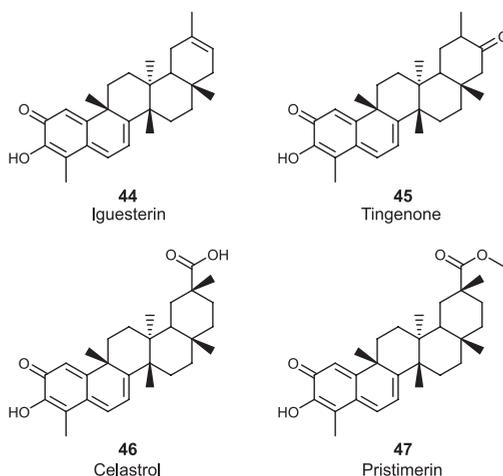


Fig. 11. Chemical structures of quinone-methide triterpenes isolated from *T. regelii*.

purification yielded phenolic phytochemicals with excellent PLpro inhibitory activities, isobavachalcone (**41**) and psoralidin (**42**) being the most potent inhibitors with  $IC_{50}$  values of 7.3 and 4.2  $\mu\text{mol L}^{-1}$ , resp. (Fig. 10). In another study reported by J.-Y. Park and colleagues (**43**) hirsutenone (**43**), isolated from *Alnus japonica*, a diarylheptanoid, showed the strongest inhibition of PLpro with an  $IC_{50}$  value of 4.1  $\mu\text{mol L}^{-1}$ . The presence of catechol and  $\alpha,\beta$ -unsaturated carbonyl moiety was found to be critical for its inhibitory potency (Fig. 10).

Furthermore, among the isolated compounds of *Tritergium regelii* as shown in Table I, iguesterin (**44**) ( $IC_{50} = 2.6 \mu\text{mol L}^{-1}$ ) was identified as a superior inhibitor of anti-SARS-CoV 3CLpro compared to quinone-methide triterpenes [tingenone (**45**) with  $IC_{50} = 9.9 \mu\text{mol}$

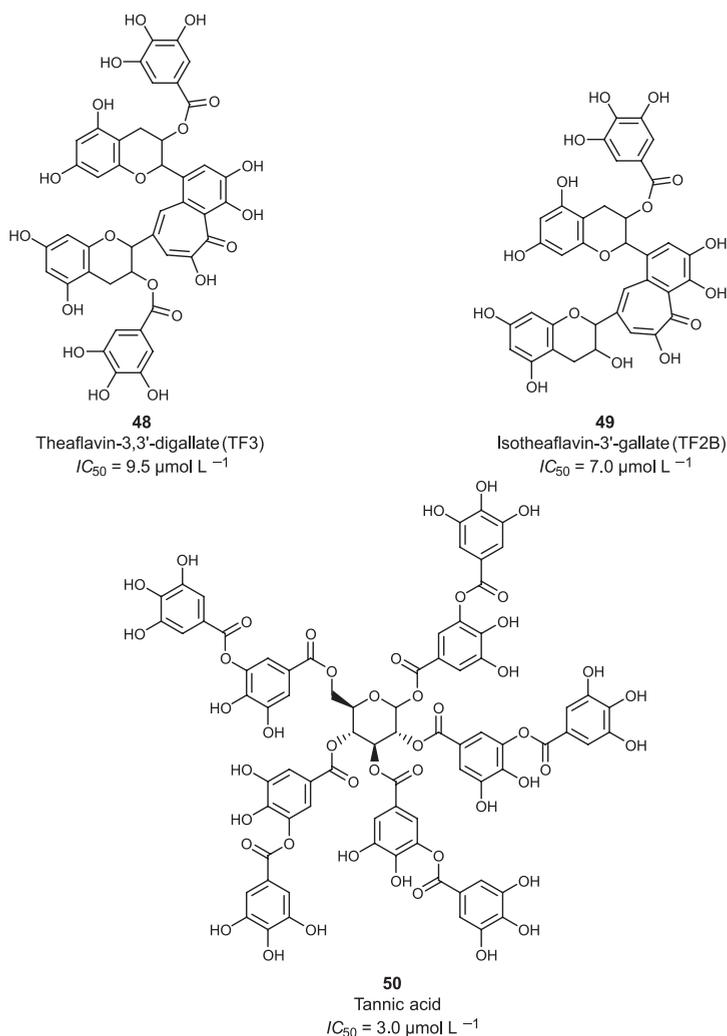


Fig. 12. Chemical structure of TF3, TF2B and tannic acid isolated from black tea extract.

L<sup>-1</sup> and celasterol (**46**) with IC<sub>50</sub> = 10.3 μmol L<sup>-1</sup>] (Fig. 11). Pristimerin (**47**) substituted with methyl ester group inhibited SARS-CoV 3CLpro activity which was two-fold greater in potency (IC<sub>50</sub> = 5.5 μmol L<sup>-1</sup>) than celasterol (**46**).

The existing literature has shown that green and black tea constitutes 20 and 78 %, resp., of global tea consumption, whereas, approximately 2 % is consumed as oolong tea (70, 71). Interestingly, these teas are from the same plant species, namely *Camellia sinensis*. These tea types also differ based on the variety of *Camellia sinensis* used in their production. For instance, green teas are made from smaller young leaves and leaf buds (*Camellia sinensis* var. *sinensis*), while black, oolong, and Pu-erh teas are made from broad leaves (*Camellia sinensis* var. *Assamica*). In addition, the crucial factor that affects the production of a particular tea is oxidation, and the process begins from leaf picking to dryness, wilting, rolling, treating and preserving. The polyphenolic content in these varieties confers a broad spectrum of biological activities including antimicrobial, antifungal, antitoxin, antioxidant and antiviral (72–75). Accordingly, Chen *et al.* (45) explored four different varieties of tea, *viz.*, green, puer, oolong and black tea, against SARS-CoV 3CLpro. Along with theaflavin (TF1), a mixture of theaflavin-3'-gallate (TF2b) and theaflavin-3-gallate (TF2a), three polyphenol compounds – theaflavin-3,3'-digallate (TF3) (**48**), isotheaflavin-3'-gallate (TF2B) (**49**) and tannic acid (**50**) were abundantly identified in the extract of black tea as effective anti-SARS-CoV inhibitors (IC<sub>50</sub> = 9.5, 7.0 and 3.0 μmol L<sup>-1</sup>, resp.) (Fig. 12). Notably,

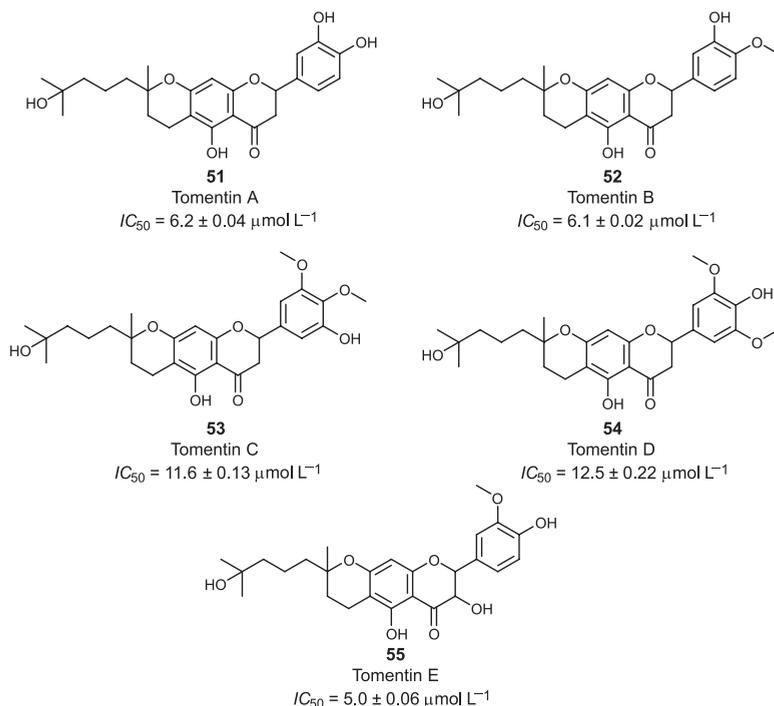


Fig. 13. Chemical structures of isolated flavonoids with anti-SARS-COV activity from the fruits of *Paulownia tomentosa*.

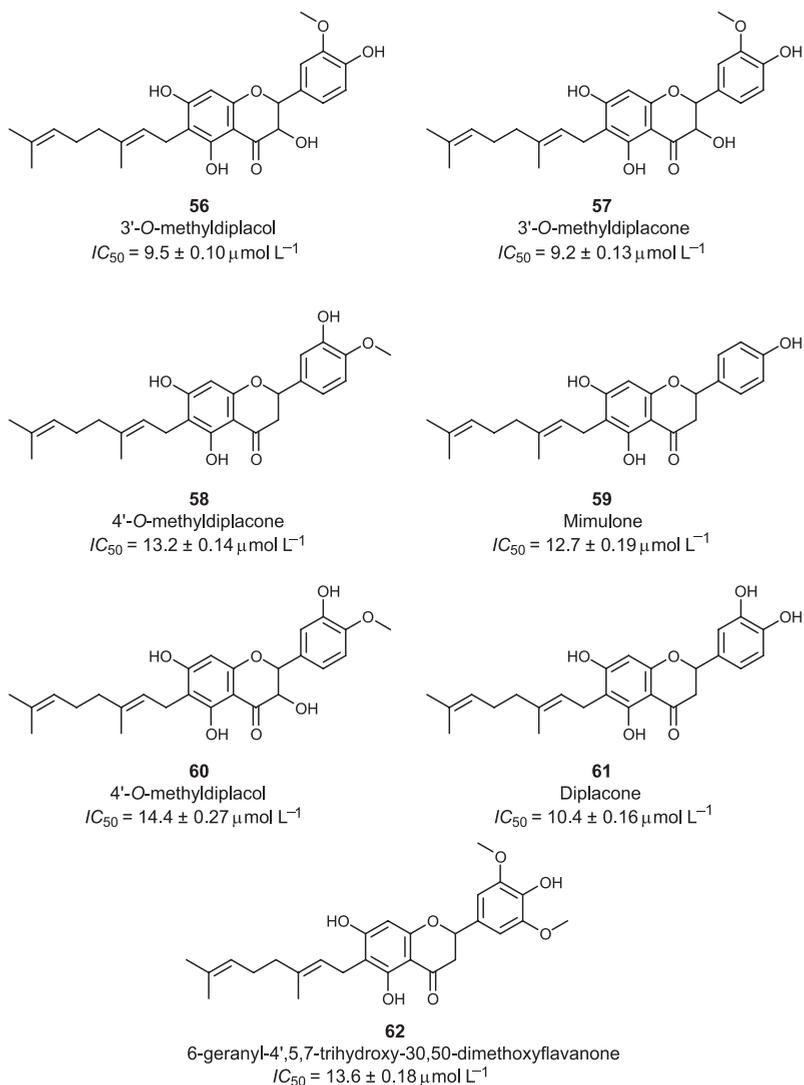


Fig. 13. Continued.

it will be very interesting to explore whether drinking black tea can be used to prevent or treat COVID-19 infection since both SARS-CoV and SARS-CoV-2 are known to dynamically replicate in the gastrointestinal tract.

A similar study evaluated twelve polyhydroxy compounds isolated from *Paulownia tomentosa* against SARS-CoV PLpro (Fig. 13) (47). Notably, compounds with a 3,4-dihydro-2H-pyran motif [tomentin A (51), tomentin B (52) and tomentin E (55)] with  $IC_{50}$  values ranging from 5.0–6.3  $\mu\text{mol L}^{-1}$  were more effective PLpro inhibitors than other iso-

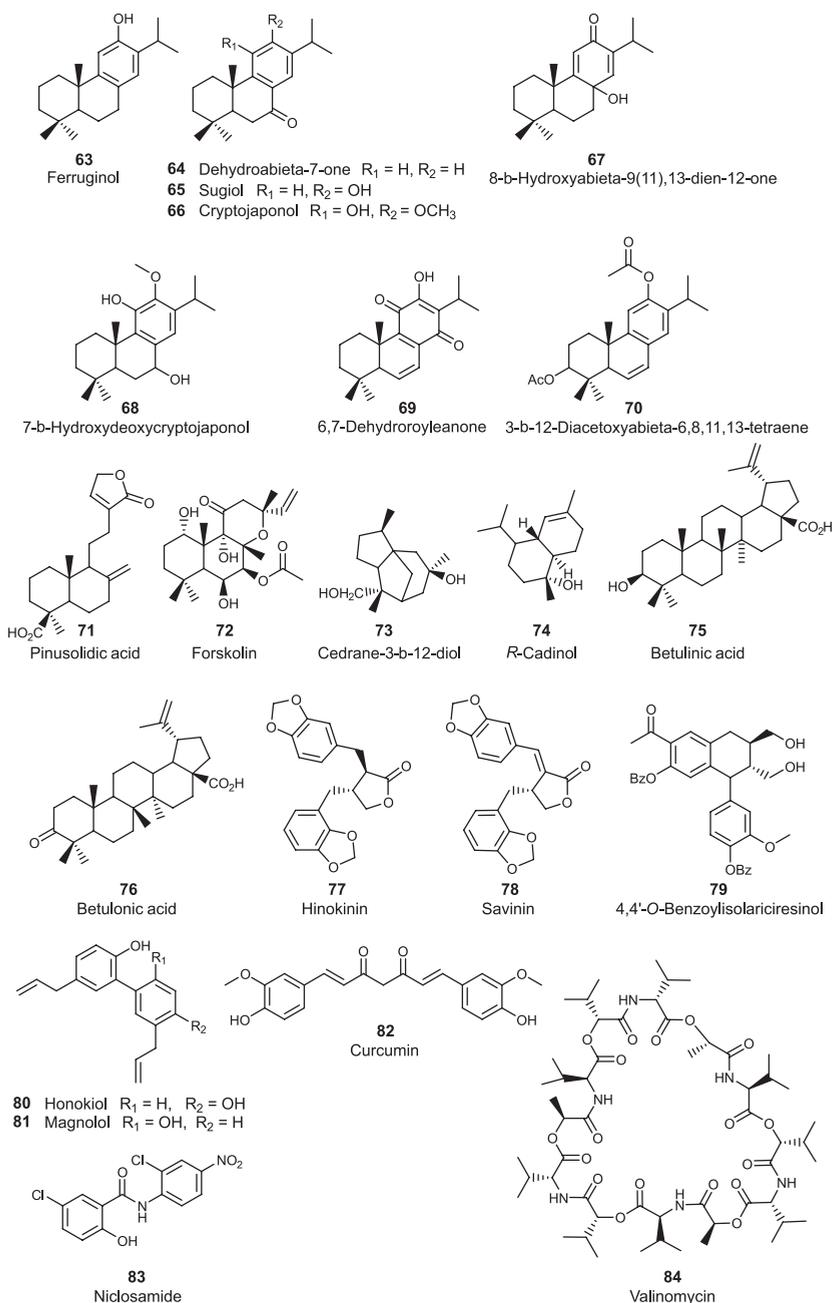


Fig. 14. Chemical structures of diterpenoids 63–72, sesquiterpenoids 73 and 74, triterpenoids 75 and 76 lignoids 77–81, phenolic compound (curcumin, 82) and two positive controls, niclosamide (83) and valinomycin (84).

lated compounds, namely, tomentin C (53), tomentin D (54), 3'-O-methyldiplacol (56), 3'-O-methyldiplacone (57), 4'-O-methyldiplacone (58), mimulone (59), diplacone (61), and 6-geranyl-4',5,7-trihydroxy-3',5'-dimethoxyflavanone (62) with  $IC_{50}$  values ranging from 9.2–14.4  $\mu\text{mol L}^{-1}$ .

Wen *et al.* (46) reported the efficacy of *Cibotium barometz* and *Dioscorea batatas* extracts against SARS-CoV 3CLpro at concentrations between 25 and 200  $\mu\text{g mL}^{-1}$  (Table I). Methanolic extracts of these plants displayed significant inhibitory potencies with 50 %-inhibitory values of 39 and 44  $\mu\text{g mL}^{-1}$ , resp. Moreover, the anti-SARS-CoV efficacy of *Cibotium barometz* and *Dioscorea batatas* extracts was superior compared to *Isatis indigotica*, *Torreya nucifera* and tea extract with 50 %-inhibitory values of 53.8, 100 and 125  $\mu\text{g mL}^{-1}$ , resp.

Wen and colleagues (46) examined the anti-SAR CoV activities of 221 phytochemicals by exploring cell-based assay to determine the SARS CoV-induced cytopathogenic outcome on Vero E6 cells. As shown in Fig. 14, twenty out of the tested compounds emerged as potent anti-SARS CoV agents at concentrations between 3.3 and 10  $\mu\text{mol L}^{-1}$ , including ten diterpenoids, namely, ferruginol (63) dehydroabieta-7-one (64), sugiol (65), cryptojaponol (66), 8- $\beta$ -hydroxyabieta-9(11),13-dien-12-one (67), 7- $\beta$ -hydroxydeoxycryptojaponol (68), 6,7-dehydroroyleanone (69), 3- $\beta$ -12-diacetoxyabieta-6,8,11,13-tetraene (70), pinusolidic acid (71), forskolin (72), two sesquiterpenoids, namely, cedrane-3- $\beta$ ,12-diol (73) and *R*-cadinol (74), two triterpenoids [betulinic acid (75) and betulonic acid (76)], five lignoids [hinokinin (77), savinin (78), 4,4'-*O*-benzoylisolariciresinol (79), honokiol (80), magnolol (81)], phenolic compound, curcumin (82), whereas niclosamide (83) and valinomycin were used as the reference compounds. Further, the 22 compounds were evaluated in a 3CL protease inhibition assay to identify the probable sites on the virus targeted by the specific anti-SARS CoV compounds using quenched fluorescence energy transfer (FRET) method. The results showed that diterpenoids (63–72) lacked SARS-CoV 3CL protease inhibition at concentrations less than 100  $\mu\text{mol L}^{-1}$ . Betulinic acid (75), savinin (78), curcumin (82) and niclosamide (83) showed the highest inhibitory activity on 3CL protease with  $IC_{50}$  values of 10, 25, 40 and 40  $\mu\text{mol L}^{-1}$ , resp., whereas betulonic acid (76) and hinokinin (77) [analogues to betulinic acid (75) and savinin (78)], resp., inhibited 3CL-protease activity with  $IC_{50}$  values > 100  $\mu\text{mol L}^{-1}$ . Hence, savinin, a lignoid purified from ethyl acetate extracts of the heartwood of *Chamaecyparis obtuse* var. *formosanal*, and betulinic acid emerged as the most potent anti-SARS-CoV compounds ( $IC_{50}$  = 25 and 10  $\mu\text{mol L}^{-1}$ , resp.) (25). The inhibitory potential of savinin for 3CLpro of SARS-CoV-2 is conceived by the presence of benzo[1,3]dioxole moiety.

## CONCLUSIONS

The present study unveils plant extracts with potent inhibitory activities against SARS-CoV. More importantly, the literature analysis revealed that the fraction RH121 of *Rheum palmatum* L. with  $IC_{50}$  = 13.76  $\pm$  0.03  $\mu\text{g mL}^{-1}$ , along with compounds isolated from other plants, such as terrestrimine isolated from *Tribulus terrestris*, cryptotanshinone, tanshinone IIA and dihydrotanshinone I (*Salvia miltiorrhiza*), xanthoangelol E (*Angelica keiskei*), papyriflavonol A (*Broussonetia papyrifera*), psoralidin (*Psoralea corylifolia*), hirsutenone (*Ahhus japonica*), tannic acid (*Camellia sinensis* var. *assamica*) and tomentin E (*Paulownia tomentosa*) with  $IC_{50}$  values ranging from 0.6–5  $\mu\text{mol L}^{-1}$  were excellent candidates for

anti-SARS-CoV targeting PLpro. Meanwhile, iguesterin with an  $IC_{50}$  value of  $2.6 \pm 0.6 \mu\text{mol L}^{-1}$  emerged as the most potent anti-SARS-CoV targeting 3CLpro.

According to all the extracted data, phytotherapy has offered a large and encouraging concept to new, safe and effective anti-SARS-CoV-2 agents. Consequently, the inhibitory potency of these medicinal plants yearns for large-scale research and development to validate their efficacy and safety for combating emerging coronavirus diseases. We also hope these findings will motivate researchers to explore the structural architecture of these compounds for the discovery of new antiviral drugs against SARS-CoVs.

*Acknowledgments.* – O. E. and O. B. conducted the search approach to avoid missing articles. Two authors, O. E. and R. O. separately assessed the full text. They would also like to thank the Mangosuthu University of Technology (MUT) for financial assistance. The authors thank the Editor-in-Chief for her professional comments and suggestions, which significantly improved quality of the paper.

*Abbreviations, acronyms, codes.* – ARDS – acute respiratory distress syndrome, 3CLpro – 3C-like protease, COVID-19 – coronavirus disease 2019, ES – effect sizes,  $IC_{50}$  – half maximal inhibitory concentration, ISG – interferon stimulated gene, MERS – Middle East respiratory syndrome, PLpro – papain-like cysteine protease, PRISMA – preferred reporting items for systematic reviews and meta-analyses, SARS-CoV – severe acute respiratory syndrome coronavirus, TF2B – isothaflavin-3'-gallate, TF3 – theaflavin-3,3'-digallate

## REFERENCES

1. F. Wu, S. Zhao, B. Yu, Y. M. Chen, W. Wang, Z. G. Song, Y. Hu, Z. W. Tao, J. H. Tian, Y. Y. Pei, M. L. Yuan, Y. L. Zhang, F. H. Dai, Y. Liu, Q. M. Wang, J. J. Zheng, L. Xu, E. C. Holmes and Y. Z. Zhang, A new coronavirus associated with human respiratory disease in China, *Nature* **579** (2020) 265–269; <https://doi.org/10.1038/s41586-020-2008-3>
2. X. Xu, P. Chen, J. Wang, J. Feng, H. Zhou, X. Li, W. Zhong and P. Hao, Evolution of the novel coronavirus from the ongoing Wuhan outbreak and modeling of its spike protein for risk of human transmission, *Sci. China Life Sci.* **63** (2020) 457–460; <https://doi.org/10.1007/s11427-020-1637-5>
3. A. J. Rodríguez-Morales, K. MacGregor, S. Kanagarajah, D. Patel and P. Schlagenhauf, Going global – Travel and the 2019 novel coronavirus, *Travel Med. Infect. Dis.* **33** (2020) 101578; <https://doi.org/10.1016/j.tmaid.2020.101578>
4. C. Huang, Y. Wang, X. Li, L. Ren, J. Zhao, Y. Hu, L. Zhang, G. Fan, J. Xu, X. Gu, Z. Cheng, T. Yu, J. Xia, Y. Wei, W. Wu, X. Xie, W. Yin, H. Li, M. Liu, Y. Xiao, H. Gao, L. Guo, J. Xie, G. Wang, R. Jiang, Z. Gao, Q. Jin, J. Wang and B. Cao, Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China, *Lancet* **395** (2020) 497–506; <https://doi.org/10.1016/S0140-6736>
5. E. Prompetchara, C. Ketloy and T. Palaga, Immune responses in COVID-19 and potential vaccines: Lessons learned from SARS and MERS epidemic, *Asian Pac. J. Allergy Immunol.* **38** (2020) 1–9; <https://doi.org/10.12932/AP-200220-0772>
6. X. Yang, Y. Yu, J. Xu, H. Shu, J. Xia, H. Liu, Y. Wu, L. Zhang, Z. Yu, M. Fang, T. Yu, Y. Wang, S. Pan, X. Zou, S. Yuan and Y. Shang, Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: A single-centered, retrospective, observational study, *Lancet Respir. Med.* **8** (2020) 475–481; [https://doi.org/10.1016/S2213-2600\(20\)30079-5](https://doi.org/10.1016/S2213-2600(20)30079-5)
7. F. Zhou, T. Yu, R. Du, G. Fan, Y. Liu, Z. Liu, J. Xiang, Y. Wang, B. Song, X. Gu, L. Guan, Y. Wei, H. Li, X. Wu, J. Xu, S. Tu, Y. Zhang, H. Chen and B. Cao, Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: A retrospective cohort study, *Lancet* **395** (2020) 1054–1062; [https://doi.org/10.1016/S0140-6736\(20\)30566-3](https://doi.org/10.1016/S0140-6736(20)30566-3)
8. Coronaviridae Study Group of the International Committee on Taxonomy of Viruses, The species severe acute respiratory syndrome-related coronavirus: Classifying 2019-nCoV and naming it SARS-CoV-2, *Nat. Microbiol.* **5** (2020) 536–544; <https://doi.org/10.1038/s41564-020-0695-z>

9. J. S. Rest and D. P. Mindell, SARS associated coronavirus has a recombinant polymerase and coronaviruses have a history of host-shifting, *Infect. Genet. Evol.* **3** (2003) 219–225; <https://doi.org/10.1016/j.meegid.2003.08.001>
10. C. C. Hon, T. Y. Lam, Z. L. Shi, A. J. Drummond, C. W. Yip, F. Zeng, P. Y. Lam and F. C. Leung, Evidence of the recombinant origin of a bat severe acute respiratory syndrome (SARS)-like coronavirus and its implications on the direct ancestor of SARS coronavirus, *J. Virol.* **82** (2008) 1819–1826; <https://doi.org/10.1128/JVI.01926-07>
11. D. Schoeman and B. C. Fielding, Coronavirus envelope protein: current knowledge, *Virol. J.* **16** (2019) Article ID 69 (22 pages); <https://doi.org/10.1186/s12985-019-1182-0>
12. P. Zhou, X. L. Yang, X. G. Wang, B. Hu, L. Zhang, W. Zhang, H. R. Si, Y. Zhu, B. Li, C. L. Huang, H. D. Chen, J. Chen, Y. Luo, H. Guo, R. D. Jiang, M. Q. Liu, Y. Chen, X. R. Shen, X. Wang, X. S. Zheng, K. Zhao, Q. J. Chen, F. Deng, L. L. Liu, B. Yan, F. X. Zhan, Y. Y. Wang, G. F. Xiao and Z. L. Shi, A pneumonia outbreak associated with a new coronavirus of probable bat origin, *Nature* **579** (2020) 270–273; <https://doi.org/10.1038/s41586-020-2012-7>
13. R. Lu, X. Zhao, J. Li, P. Niu, B. Yang, H. Wu, W. Wang, H. Song, B. Huang, N. Zhu, Y. Bi, X. Ma, F. Zhan, L. Wang, T. Hu, H. Zhou, Z. Hu, W. Zhou, L. Zhao, J. Chen, Y. Meng, J. Wang, Y. Lin, J. Yuan, Z. Xie, J. Ma, W. J. Liu, D. Wang, W. Xu, E. C. Holmes, G. F. Gao, G. Wu, W. Chen, W. Shi and W. Tan, Genomic characterisation and epidemiology of 2019 novel coronavirus: Implications for virus origins and receptor binding, *Lancet* **395** (2020) 565–574; [https://doi.org/10.1016/S0140-6736\(20\)30251-8](https://doi.org/10.1016/S0140-6736(20)30251-8)
14. K. Anand, J. Ziebuhr, P. Wadhvani, J. R. Mesters and R. Hilgenfeld, Coronavirus main proteinase (3CLpro) structure: Basis for design of anti-SARS drugs, *Science* **300** (2003) 1763–1767; <https://doi.org/10.1126/science.1085658>
15. M. T. ul Qamar, S. M. Alqahtani, M. A. Alamri and L. L. Chen, Structural basis of SARS-CoV-2 3CLpro and anti-COVID-19 drug discovery from medicinal plants, *J. Pharm. Anal.* **10** (2020) 313–319; <https://doi.org/10.1016/j.jpha.2020.03.009>
16. A. L. Totura and R. S. Baric, SARS coronavirus pathogenesis: Host innate immune responses and viral antagonism of interferon, *Curr. Opin. Virol.* **3** (2012) 264–275; <https://doi.org/10.1016/j.coviro.2012.04.004>
17. T. H. Mogensen and S. R. Paludan, Molecular pathways in virus-induced cytokine production, *Microbiol. Mol. Biol. Rev.* **65** (2001) 131–150; <https://doi.org/10.1128/MMBR.65.1.131-150.2001>
18. C. K. Wong, C. W. Lam, A. K. Wu, W. K. Ip, N. L. Lee, I. H. Chan, L. C. Lit, D. S. Hui, M. H. Chan, S. S. Chung and J. J. Sung, Plasma inflammatory cytokines and chemokines in severe acute respiratory syndrome, *Clin. Exp. Immunol.* **136** (2004) 95–103; <https://doi.org/10.1111/j.1365-2249.2004.02415.x>
19. O. Ebenezer, M. A. Jordaan, R. E. Ogunsakin and M. Shapi, Potential SARS-COV preclinical (in vivo) compounds targeting COVID-19 main protease: A meta-analysis and molecular docking studies, *Hippokratia* **24** (2020) 99–106.
20. K. Van Reeth, S. Van Gucht and M. Pensaert, In vivo studies on cytokine involvement during acute viral respiratory disease of swine: Troublesome but rewarding, *Vet. Immunol. Immunopathol.* **87** (2002) 161–168; [https://doi.org/10.1016/s0165-2427\(02\)00047-8](https://doi.org/10.1016/s0165-2427(02)00047-8)
21. C. Y. Cheung, L. L. Poon, A. S. Lau, W. Luk, Y. L. Lau, K. F. Shortridge, S. Gordon, Y. Guan and J. S. Peiris, Induction of proinflammatory cytokines in human macrophages by influenza A (H5N1) viruses: A mechanism for the unusual severity of human disease?, *Lancet* **360** (2002) 1831–1837; [https://doi.org/10.1016/s0140-6736\(02\)11772-7](https://doi.org/10.1016/s0140-6736(02)11772-7)
22. L. Zhang, D. Lin, X. Sun, U. Curth, C. Drosten, L. Sauerhering, S. Becker, X. Rox and R. Hilgenfeld, Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved  $\alpha$ -ketoamide inhibitors, *Science* **368** (2020) 409–412; <https://doi.org/10.1126/science.abb3405>
23. R. Hilgenfeld, From SARS to MERS: Crystallographic studies on coronaviral proteases enable antiviral drug design, *FEBS J.* **281** (2014) 4085–4096; <https://doi.org/10.1111/febs.12936>
24. S. Y. Li, C. Chen, H. Q. Zhang, H. Y. Guo, H. Wang, L. Wang, X. Zhang, S. N. Hua, J. Yu, P. G. Xiao, R. S. Li and X. Tan, Identification of natural compounds with antiviral activities against SARS-associated coronavirus, *Antiviral. Res.* **67** (2005) 18–23; <https://doi.org/10.1016/j.antiviral.2005.02.007>

25. C. C. Wen, Y. H. Kuo, J. T. Jan, P. H. Liang, S. Y. Wang, H. G. Liu, C. K. Lee, S. T. Chang, C. J. Kuo, S. S. Lee, C. C. Hou, P. W. Hsiao, S. C. Chien, L. F. Shyur and N. S. Yang, Specific plant terpenoids and lignoids possess potent antiviral activities against severe acute respiratory syndrome coronavirus, *J. Med. Chem.* **50** (2007) 4087–4095; <https://doi.org/10.1021/jm070295s>
26. L. T. Lin, W. C. Hsu and C. C. Lin, Antiviral natural products and herbal medicines, *J. Tradit. Complement. Med.* **4** (2014) 24–35; <https://doi.org/10.4103/2225-4110.124335>
27. T. Pillaiyar, M. Manickam, V. Namasivayam, Y. Hayashi and S. H. Jung, An Overview of severe acute respiratory syndrome-Coronavirus (SARS-CoV) 3CL protease inhibitors: Peptidomimetics and small molecule chemotherapy, *J. Med. Chem.* **59** (2016) 6595–6628; <https://doi.org/10.1021/acs.jmedchem.5b01461>
28. L. Shamseer, D. Moher, M. Clarke, D. Ghersi, A. Liberati, M. Petticrew, P. Shekelle and L. A. Stewart, (PRISMA-P group), Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015: Elaboration and explanation, *BMJ* **350** (2015) Article ID g7647; <https://doi.org/10.1136/bmj.g7647>
29. W. Yan, K. Ohtani, R. Kasai and K. Yamasaki, Steroidal saponins from fruits of *Tribulus terrestris*, *Phytochemistry* **42** (1996) 1417–1422; [https://doi.org/10.1016/0031-9422\(96\)00131-8](https://doi.org/10.1016/0031-9422(96)00131-8)
30. I. Kostova and D. Dinchev, Saponins in *Tribulus terrestris* – Chemistry and bioactivity, *Phytochem. Rev.* **4** (2005) 111–137; <https://doi.org/10.1007/s11101-005-2833-x>
31. N. Mulinacci, P. Vignolini, G. la Marca, G. Pieraccini, M. Innocenti and F. F. Vincieri, Food supplements of *Tribulus terrestris* L.: An HPLC-ESI-MS method for an estimation of the saponin content, *Chromatographia* **57** (2003) 581–592; <https://doi.org/10.1007/BF02491733>
32. W. Zheng, F. Wang, Y. Zhao, X. Sun, L. Kang, Z. Fan, L. Qiao, R. Yan, S. Liu and B. Ma, Rapid characterization of constituents in *Tribulus terrestris* from different habitats by UHPLC/Q-TOF MS, *J. Am. Soc. Mass Spectrom.* **28** (2017) 2302–2318; <https://doi.org/10.1007/s13361-017-1761-5>
33. C. Tian, Z. Zhang, H. Wang, Y. Guo, J. Zhao and M. Liu, Extraction technology, component analysis, and in vitro antioxidant and antibacterial activities of total flavonoids and fatty acids from *Tribulus terrestris* L. fruits, *Biomed. Chromatogr.* **33** (2019) e4474; <https://doi.org/10.1002/bmc.4474>
34. J. Kaushik, S. Tandon, R. Bhardwaj, T. Kaur, S. K. Singla, J. Kumar and C. Tandon, Delving into the antiuro lithiatic potential of *Tribulus terrestris* extract through – In vivo efficacy and preclinical safety investigations in Wistar rats, *Sci. Rep.* **9** (2019) Article ID 15969 (13 pages); <https://doi.org/10.1038/s41598-019-52398-w>
35. Y. H. Song, D. W. Kim, M. J. Curtis-Long, H. J. Yuk, Y. Wang, N. Zhuang, K. H. Lee, K. S. Jeon and K. H. Park, Papain-like protease (PLpro) inhibitory effects of cinnamic amides from *Tribulus terrestris* fruits, *Biol. Pharm. Bull.* **37** (2014) 1021–1028; <https://doi.org/10.1248/bpb.b14-00026>
36. C. W. Lin, F. J. Tsai, C. H. Tsai, C. C. Lai, L. Wan, T. Y. Ho, C. C. Hsieh and P. D. Chao, Anti-SARS coronavirus 3C-like protease effects of *Isatis indigotica* root and plant-derived phenolic compounds, *Antiviral Res.* **68** (2005) 36–42; <https://doi.org/10.1016/j.antiviral.2005.07.002>
37. J. Y. Park, J. H. Kim, Y. M. Kim, H. J. Jeong, D. W. Kim, K. H. Park, H. J. Kwon, S. J. Park, W. S. Lee and Y. B. Ryu, Tanshinones as selective and slow-binding inhibitors for SARS-CoV cysteine proteases, *Bioorg. Med. Chem.* **20** (2012) 5928–5935; <https://doi.org/10.1016/j.bmc.2012.07.038>
38. W. Luo, X. Su, S. Gong, Y. Qin, W. Liu, J. Li, H. Yu and Q. Xu, Anti-SARS coronavirus 3C-like protease effects of *Rheum palmatum* L. extracts, *Biosci. Trends* **3** (2009) 124–126.
39. J. Y. Park, J. A. Ko, D. W. Kim, Y. M. Kim, H. J. Kwon, H. J. Jeong, C. Y. Kim, K. H. Park, W. S. Lee and Y. B. Ryu, Chalcones isolated from *Angelica keiskei* inhibit cysteine proteases of SARS-CoV, *J. Enzyme Inhib. Med. Chem.* **31** (2016) 23–30; <https://doi.org/10.3109/14756366.2014.1003215>
40. J. Y. Park, H. J. Yuk, H. W. Ryu, S. H. Lim, K. S. Kim, K. H. Park, Y. B. Ryu and W. S. Lee, Evaluation of polyphenols from *Broussonetia papyrifera* as coronavirus protease inhibitors, *J. Enzyme Inhib. Med. Chem.* **32** (2017) 504–515; <http://doi.org/10.1080/14756366.2016.1265519>

41. Y. B. Ryu, H. J. Jeong, J. H. Kim, Y. M. Kim, J. Y. Park, D. Kim, T. T. Nguyen, S. J. Park, J. S. Chang, K. H. Park, M. C. Rho and W. S. Lee, Biflavonoids from *Torreya nucifera* displaying SARS-CoV 3CL(pro) inhibition, *Bioorg. Med. Chem.* **18** (2010) 7940–7947; <https://doi.org/10.1016/j.bmc.2010.09.035>
42. D. W. Kim, K. H. Seo, M. J. Curtis-Long, K. Y. Oh, J. W. Oh, J. K. Cho, K. H. Lee and K. H. Park, Phenolic phytochemical displaying SARS-CoV papain-like protease inhibition from the seeds of *Psoralea corylifolia*, *J. Enzyme Inhib. Med. Chem.* **29** (2014) 59–63; <https://doi.org/10.3109/14756366.2012.753591>
43. J.-Y. Park, H. J. Jeong, J. H. Kim, Y. M. Kim, S.-J. Park, D. Kim, K. H. Park, W. S. Lee and Y. B. Ryu, Diarylheptanoids from *Alnus japonica* inhibit papain-like protease of severe acute respiratory syndrome coronavirus, *Biol. Pharm. Bull.* **35** (2012) 2036–2042; <https://doi.org/10.1248/bpb.b12-00623>
44. B. Ryu, J. Park, M. Kim, Y. Lee, D. Seo, S. Chang, H. Park, C. Rho and S. Lee, SARS-CoV 3CLpro inhibitory effects of quinone-methide triterpenes from *Tripterygium regelii*, *Bioorg. Med. Chem. Lett.* **20** (2010) 1873–1876; <https://doi.org/10.1016/j.bmcl.2010.01.152>
45. C. N. Chen, C. P. Lin, K. K. Huang, W. C. Chen, H. P. Hsieh, P. H. Liang and J. T. Hsu, Inhibition of SARS-CoV 3C-like protease activity by theaflavin-3,3'-digallate (TF3), *Evid. Based Complement. Alternat. Med.* **2** (2005) 209–215; <https://doi.org/10.1093/ecam/neh081>
46. C. C. Wen, L. F. Shyur, J. T. Jan, P. H. Liang, C. J. Kuo, P. Arulsevan, J. B. Wu, S. C. Kuo and N. S. Yang, Traditional Chinese medicine herbal extracts of *Cibotium barometz*, *Gentiana scabra*, *Dioscorea batatas*, *Cassia tora*, and *Taxillus chinensis* inhibit SARS-CoV replication, *J. Tradit. Complement. Med.* **1** (2011) 41–50; [https://doi.org/10.1016/s2225-4110\(16\)30055-4](https://doi.org/10.1016/s2225-4110(16)30055-4)
47. J. K. Cho, M. J. Curtis-Long, K. H. Lee, D. W. Kim, H. W. Ryu, H. J. Yuk and K. H. Park, Geranylated flavonoids displaying SARS-CoV papain-like protease inhibition from the fruits of *Paulownia tomentosa*, *Bioorg. Med. Chem.* **21** (2013) 3051–3057; <https://doi.org/10.1016/j.bmc.2013.03.027>
48. G. W. Qin and R. S. Xu. Recent advances on bioactive natural products from Chinese medicinal plants, *Med. Res. Rev.* **18** (1998) 375–382; [https://doi.org/10.1002/\(sici\)1098-1128\(199811](https://doi.org/10.1002/(sici)1098-1128(199811)
49. X. Wu, G. Qin, K. K. Cheung and K. F. Cheng, New alkaloids from *Isatis indigotica*, *Tetrahedron* **53** (1997) 13323–13328.
50. C. Jie, Z. Luo, H. Chen, M. Wang, C. Yan, Z. F. Mao, G. K. Xiao, H. Kurihara, Y. F. Li and R. R. He, Indirubin, a bisindole alkaloid from *Isatis indigotica*, reduces H1N1 susceptibility in stressed mice by regulating MAVS signaling, *Oncotarget* **8** (2017) 105615–105629; <https://doi.org/10.18632/oncotarget.22350>
51. S. L. Hsuan, S. C. Chang, S. Y. Wang, T. L. Liao, T. T. Jong, M. S. Chien, W. C. Lee, S. S. Chen and J. W. Liao, The cytotoxicity to leukemia cells and antiviral effects of *Isatis indigotica* extracts on pseudorabies virus, *J. Ethnopharmacol.* **123** (2009) 61–67; <https://doi.org/10.1016/j.jep.2009.02.028>
52. M. Iranshahi, R. Rezaee, H. Parhiz, A. Roohbakhsh and F. Soltani, Protective effects of flavonoids against microbes and toxins: The cases of hesperidin and hesperetin, *Life Sci.* **137** (2015) 125–132; <https://doi.org/10.1016/j.lfs.2015.07.014>
53. T. N. Kaul, E. Middleton, Jr. and P. L. Ogra, Antiviral effect of flavonoids on human viruses, *J. Med. Virol.* **15** (1985) 71–79; <https://doi.org/10.1002/jmv.1890150110>
54. W. S. Lee, J. R. Kim, J. M. Han, K. C. Jang, D. E. Sok and T. S. Jeong, Antioxidant activities of abietane diterpenoids isolated from *Torreya nucifera* leaves, *J. Agric. Food Chem.* **54** (2006) 5369–5374; <https://doi.org/10.1021/jf060896c>
55. K. Bae, *The Medicinal Plants of Korea*, Kyo-Hak Publishing Co., Seoul 2000, pp. 260.
56. J. Oh, H. S. Rho, Y. Yang, J. Y. Yoon, J. Lee, Y. D. Hong, H. C. Kim, S. S. Choi, T. W. Kim, S. S. Shin and J. Y. Cho, Extracellular signal-regulated kinase is a direct target of the anti-inflammatory compound ameto flavone derived from *Torreya nucifera*, *Mediators Inflamm.* **2013** (2013) Article ID 761506 (11 pages); <https://doi.org/10.1155/2013/761506>
57. S. P. Chen, M. Dong, K. Kita, Q. W. Shi, B. Cong, W. Z. Guo, S. Sugaya, K. Sugita and N. Suzuki, Anti-proliferative and apoptosis-inducible activity of labdane and abietane diterpenoids from the pulp of *Torreya nucifera* in HeLa cells, *Mol. Med. Rep.* **3** (2010) 673–678; [https://doi.org/10.3892/mmr\\_00000315](https://doi.org/10.3892/mmr_00000315)

58. S. H. Kim, J. G. Park, Y. D. Hong, E. Kim, K. S. Baik, D. H. Yoon, S. Kim, M. N. Lee, H. S. Rho, S. S. Shin and J. Y. Cho, Src/Syk/IRAK1-targeted anti-inflammatory action of *Torreya nucifera* butanol fraction in lipopolysaccharide-activated RAW264.7 cells, *J. Ethnopharmacol.* **188** (2016) 167–176; <https://doi.org/10.1016/j.jep.2016.05.008>
59. P. G. Latha, D. A. Evans, K. R. Panikkar and K. K. Jayavardhanan, Immunomodulatory and anti-tumour properties of *Psoralea corylifolia* seeds, *Fitoterapia* **71** (2000) 223–231; [https://doi.org/10.1016/s0367-326x\(99\)00151-3](https://doi.org/10.1016/s0367-326x(99)00151-3)
60. J. P. Kotiyal and D. P. Sharma, Phytochemical studies of *Psoralea* species, *Bull. Medico-Ethnobot. Res.* **13** (1992) 209–223.
61. G. Jiangning, W. Xinchu, W. Hou, L. Qinghua and B. Kaishun, Antioxidants from a Chinese medicinal herb *Psoralea corylifolia* L., *Food Chem.* **91** (2005) 287–292.
62. G. Xiao, G. Li, L. Chen, Z. Zhang, J. J. Yin, T. Wu, Z. Cheng, X. Wei and Z. Wang, Isolation of antioxidants from *Psoralea corylifolia* fruits using high-speed counter-current chromatography guided by thin layer chromatography-antioxidant autographic assay, *J. Chromatogr. A* **1217** (2010) 5470–5476; <https://doi.org/10.1016/j.chroma.2010.06.041>
63. S. Yin, C. Q. Fan, Y. Wang, L. Dong and J. M. Yue, Antibacterial prenylflavone derivatives from *Psoralea corylifolia*, and their structure-activity relationship study, *Bioorg. Med. Chem.* **12** (2004) 4387–4392; <https://doi.org/10.1016/j.bmc.2004.06.014>
64. N. A. Khatune, M. E. Islam, M. E. Haque, P. Khondkar and M. M. Rahman, Antibacterial compounds from the seeds of *Psoralea corylifolia*, *Fitoterapia* **75** (2004) 228–230; <https://doi.org/10.1016/j.fitote.2003.12.018>
65. S. Chanda, M. Kaneria and R. Nair, Antibacterial activity of *Psoralea corylifolia* L. seed and aerial parts with various extraction methods, *Res. J. Microbiol.* **6** (2011) 124–131; <https://doi.org/10.3923/jm.2011.124.13>
66. C. H. Chen, T. L. Hwang, L. C. Chen, T. H. Chang, C. S. Wei and J. J. Chen, Isoflavones and anti-inflammatory constituents from the fruits of *Psoralea corylifolia*, *Phytochemistry* **143** (2017) 186–193; <https://doi.org/10.1016/j.phytochem.2017.08.004>
67. L. T. Yi, Y. C. Li, Y. Pan, J. M. Li, Q. Xu, S. F. Mo, C. F. Qiao, F. X. Jiang, H. X. Xu, X. B. Lu, L. D. Kong and H. F. Kung, Antidepressant-like effects of psoralidin isolated from the seeds of *Psoralea corylifolia* in the forced swimming test in mice, *Prog. Neuropsychopharmacol. Biol. Psychiatry* **32** (2008) 510–519; <https://doi.org/10.1016/j.pnpbp.2007.10.005>
68. N. J. Sun, S. H. Woo, J. M. Cassidy and R. M. Snapka, DNA polymerase and topoisomerase II inhibitors from *Psoralea corylifolia*, *J. Nat. Prod.* **61** (1998) 362–366; <https://doi.org/10.1021/np970488q>
69. C. Cheng, S. Yu-Feng, H. Yang, L. Lei, C. Wei-Chao, W. Gao-Xue and Z. Bin, Highly efficient inhibition of spring viraemia of carp virus replication in vitro mediated by bavachin, a major constituent of *Psoralea corylifolia* Lynn., *Virus Res.* **255** (2018) 24–35; <https://doi.org/10.1016/j.virusres.2018.06.002>
70. C. Cabrera, R. Giménez and M. C. López, Determination of tea components with antioxidant activity, *J. Agric. Food Chem.* **51** (2003) 4427–4435; <https://doi.org/10.1021/jf0300801>
71. M. Weerawatanakorn, W. L. Hung, M. H. Pan, S. Li, D. Li, X. Wan and C. T. Ho, Chemistry and health beneficial effects of oolong tea and theasinensins, *Food Sci. Hum. Well.* **4** (2015) 133–146; <https://doi.org/10.1016/j.FSHW.2015.10.002>
72. W. C. Reygaert, The antimicrobial possibilities of green tea, *Front. Microbiol.* **5** (2014) Article ID 434 (8 pages); <https://doi.org/10.3389/fmicb.2014.00434>
73. M. Friedman, Overview of antibacterial, antitoxin, antiviral, and antifungal activities of tea flavonoids and teas, *Mol. Nutr. Food Res.* **51** (2007) 116–134; <https://doi.org/10.1002/mnfr.200600173>
74. J. M. Song, K. H. Lee and B. L. Seong, Antiviral effect of catechins in green tea on influenza virus, *Antiviral Res.* **68** (2005) 66–74; <https://doi.org/10.1016/j.antiviral.2005.06.010>
75. Z. F. Yang, L. P. Bai, W. B. Huang, X. Z. Li, S. S. Zhao, N. S. Zhong and Z. H. Jiang, Comparison of in vitro antiviral activity of tea polyphenols against influenza A and B viruses and structure-activity relationship analysis, *Fitoterapia* **93** (2014) 47–53; <https://doi.org/10.1016/j.fitote.2013.12.011>